# Mag-Bind® SeqDTR™

Revision No: 2.0

## Quick Guide

Please visit www.omegabiotek.com for a downloadable user manual containing additional protocols, troubleshooting tips, and ordering information.

| Product | M1300-05 | M1300-08 | M1300-50 |
|---|---|---|---|
| Mag-Bind® SeqDTR | 5 mL | 50 mL | 500 mL |
| Preparations | 500*<br>1,000** | 5,000*<br>10,000** | 50,000*<br>100,000** |

* *Based on a typical 10 µL reaction volume in a 96-well format*
** *Based on a typical 5 µL reaction volume in a 384-well format*

**Supplied by user:**

- 85% ethanol (do not use denatured ethanol)
- Magnetic separation device compatible with 96-well PCR plates
- Multichannel pipet
- Reservoirs
- 96-well or 384-well plate capable of being used in sequencers
- Elution Buffer (Cat# PDR048 or 10 mM Tris pH 8.5, TE Buffer, 0.1 mM EDTA, or di$H_2O$)

## Protocol for 96 Plates

BIND

1. Thoroughly shake the Mag-Bind® SeqDTR™ to fully resuspend the magnetic beads.
2. Add 10 µL Mag-Bind® SeqDTR™ to each well. Use 10 µL Mag-Bind® SeqDTR™ regardless of the volume of the sequencing reaction.
3. Add 85% ethanol according to table below and mix the sample thoroughly by pipetting up and down 7-10 times. Do not use denatured ethanol. Always prepare fresh 85% ethanol within 3 days of use and store tightly capped.

| Reaction volume (µL) | 85% Ethanol (µL) |
|---|---|
| 5 | 30 |
| 10 | 40 |
| 15 | 50 |
| 20 | 60 |

4. Place the plate on a magnetic separation device to magnetize the Mag-Bind® SeqDTR. Let sit at room temperature until the Mag-Bind® SeqDTR™ is completely cleared from solution. Aspirate and discard the supernatant. Do not disturb the Mag-Bind® SeqDTR™.

WASH

5. Add 100 µL 85% ethanol to each well. It is not necessary to resuspend the Mag-Bind® SeqDTR. Let sit at room temperature until the Mag-Bind® SeqDTR™ is completely cleared from solution. Aspirate and discard the supernatant. Do not disturb the Mag-Bind® SeqDTR™.
6. Repeat Steps 5 for a second 85% ethanol wash step.
7. Leave the plate on the magnetic separation device for 10-15 minutes to air dry the Mag-Bind® SeqDTR™. Remove any residue liquid with a pipettor. It is important to dry the Mag-Bind® SeqDTR™ before elution. Residual ethanol may interfere with downstream applications.

ELUTE

8. Add 40 µL Elution Buffer (or 10 mM Tris pH 8.5, TE Buffer, 0.1 mM EDTA, or di$H_2O$) to each well. Pipet up and down 20 times to mix thoroughly. Let sit at room temperature for 5 minutes.
9. Place the plate on a magnetic separation device to magnetize the Mag-Bind® SeqDTR™. Let sit at room temperature until the Mag-Bind® SeqDTR™ is completely cleared from solution.
10. Transfer 30-35 µL cleared supernatant containing purified sequencing product to a new plate capable of being used in sequencer.

## Protocol for 384-well Plates

BIND

1. Thoroughly shake the Mag-Bind® SeqDTR™ to fully resuspend the magnetic beads.
2. Add 5 µL Mag-Bind® SeqDTR™ to each well. Use 5 µL Mag-Bind® SeqDTR™ regardless of the volume of the sequencing reaction.
3. Add 85% ethanol according to table below and mix the sample thoroughly by pipetting up and down 7-10 times. Do not use denatured ethanol. Always prepare fresh 85% ethanol within 3 days of use and store tightly capped.

| Reaction volume (µL) | 85% Ethanol (µL) |
|---|---|
| 5 | 14.3 |
| 10 | 21.4 |
| 15 | 28.6 |

WASH

4. Place the plate on a magnetic separation device to magnetize the Mag-Bind® SeqDTR. Let sit at room temperature until the Mag-Bind® SeqDTR is completely cleared from solution. Aspirate and discard the supernatant. Do not disturb the Mag-Bind® SeqDTR™.
5. Add 30 µL 85% ethanol to each well. It is not necessary to resuspend the Mag-Bind® SeqDTR. Let sit at room temperature until the Mag-Bind® SeqDTR is completely cleared from solution. Aspirate and discard the supernatant. Do not disturb the Mag-Bind® SeqDTR.
6. Repeat Step 5 for a second 85% ethanol wash step.
7. Leave the plate on the magnetic separation device for 10-15 minutes to air dry the Mag-Bind® SeqDTR™. Remove any residue liquid with a pipettor. It is important to dry the Mag-Bind® SeqDTR™ before elution. Residual ethanol may interfere with downstream applications.

ELUTE

8. Add 15-20 µL Elution Buffer (or 10 mM Tris pH 8.5, TE Buffer, 0.1 mM EDTA, or di$H_2O$) to each well. Pipet up and down 20 times to mix thoroughly. Let sit at room temperature for 5 minutes.
9. Place the plate on a magnetic separation device to magnetize the Mag-Bind® SeqDTR™. Let sit at room temperature until the Mag-Bind® SeqDTR™ is completely cleared from solution.
10. Transfer the cleared supernatant containing purified sequencing product to a new plate capable of being used in sequencer.