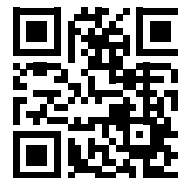


## Quick Guide

Please visit [www.omegabiotek.com](http://www.omegabiotek.com) for a downloadable user manual containing additional protocols, troubleshooting tips, and ordering information.



| Product          | M1300-05 | M1300-08 | M1300-50  |
|------------------|----------|----------|-----------|
| Mag-Bind® SeqDTR | 5 mL     | 50 mL    | 500 mL    |
| Preparations     | 500*     | 5,000*   | 50,000*   |
|                  | 1,000**  | 10,000** | 100,000** |

\* Based on a typical 10 µL reaction volume in a 96-well format

\*\* Based on a typical 5 µL reaction volume in a 384-well format

### Supplied by user:

- 85% ethanol (do not use denatured ethanol)
- Magnetic separation device compatible with 96-well PCR plates
- Multichannel pipet
- Reservoirs
- 96-well or 384-well plate capable of being used in sequencers
- Elution Buffer (Cat# PDR048 or 10 mM Tris pH 8.5, TE Buffer, 0.1 mM EDTA, or diH<sub>2</sub>O)

## Protocol for 96 Plates

1. Thoroughly shake the Mag-Bind® SeqDTR™ to fully resuspend the magnetic beads.
2. Add 10 µL Mag-Bind® SeqDTR™ to each well. Use 10 µL Mag-Bind® SeqDTR™ regardless of the volume of the sequencing reaction.
3. Add 85% ethanol according to table below and mix the sample thoroughly by pipetting up and down 7-10 times. Do not use denatured ethanol. Always prepare fresh 85% ethanol within 3 days of use and store tightly capped.

| Reaction volume (µL) | 85% Ethanol (µL) |
|----------------------|------------------|
| 5                    | 30               |
| 10                   | 40               |
| 15                   | 50               |
| 20                   | 60               |

4. Place the plate on a magnetic separation device to magnetize the Mag-Bind® SeqDTR™. Let sit at room temperature until the Mag-Bind® SeqDTR™ is completely cleared from solution. Aspirate and discard the supernatant. Do not disturb the Mag-Bind® SeqDTR™.
5. Add 100 µL 85% ethanol to each well. It is not necessary to resuspend the Mag-Bind® SeqDTR™. Let sit at room temperature until the Mag-Bind® SeqDTR™ is completely cleared from solution. Aspirate and discard the supernatant. Do not disturb the Mag-Bind® SeqDTR™.
6. Repeat Steps 5 for a second 85% ethanol wash step.
7. Leave the plate on the magnetic separation device for 10-15 minutes to air dry the Mag-Bind® SeqDTR™. Remove any residue liquid with a pipettor. It is important to dry the Mag-Bind® SeqDTR™ before elution. Residual ethanol may interfere with downstream applications.
8. Add 40 µL Elution Buffer (or 10 mM Tris pH 8.5, TE Buffer, 0.1 mM EDTA, or diH<sub>2</sub>O) to each well. Pipet up and down 20 times to mix thoroughly. Let sit at room temperature for 5 minutes.
9. Place the plate on a magnetic separation device to magnetize the Mag-Bind® SeqDTR™. Let sit at room temperature until the Mag-Bind® SeqDTR™ is completely cleared from solution.
10. Transfer 30-35 µL cleared supernatant containing purified sequencing product to a new plate capable of being used in sequencer.

BIND

WASH

ELUTE

### Protocol for 384-well Plates

1. Thoroughly shake the Mag-Bind® SeqDTR™ to fully resuspend the magnetic beads.
2. Add 5 µL Mag-Bind® SeqDTR™ to each well. Use 5 µL Mag-Bind® SeqDTR™ regardless of the volume of the sequencing reaction.
3. Add 85% ethanol according to table below and mix the sample thoroughly by pipetting up and down 7-10 times. Do not use denatured ethanol. Always prepare fresh 85% ethanol within 3 days of use and store tightly capped.

| Reaction volume (µL) | 85% Ethanol (µL) |
|----------------------|------------------|
| 5                    | 14.3             |
| 10                   | 21.4             |
| 15                   | 28.6             |

4. Place the plate on a magnetic separation device to magnetize the Mag-Bind® SeqDTR. Let sit at room temperature until the Mag-Bind® SeqDTR is completely cleared from solution. Aspirate and discard the supernatant. Do not disturb the Mag-Bind® SeqDTR™.
5. Add 30 µL 85% ethanol to each well. It is not necessary to resuspend the Mag-Bind® SeqDTR. Let sit at room temperature until the Mag-Bind® SeqDTR is completely cleared from solution. Aspirate and discard the supernatant. Do not disturb the Mag-Bind® SeqDTR.
6. Repeat Step 5 for a second 85% ethanol wash step.
7. Leave the plate on the magnetic separation device for 10-15 minutes to air dry the Mag-Bind® SeqDTR™. Remove any residue liquid with a pipettor. It is important to dry the Mag-Bind® SeqDTR™ before elution. Residual ethanol may interfere with downstream applications.
8. Add 15-20 µL Elution Buffer (or 10 mM Tris pH 8.5, TE Buffer, 0.1 mM EDTA, or diH<sub>2</sub>O) to each well. Pipet up and down 20 times to mix thoroughly. Let sit at room temperature for 5 minutes.
9. Place the plate on a magnetic separation device to magnetize the Mag-Bind® SeqDTR™. Let sit at room temperature until the Mag-Bind® SeqDTR™ is completely cleared from solution.
10. Transfer the cleared supernatant containing purified sequencing product to a new plate capable of being used in sequencer.

BIND

WASH

ELUTE