omega BIO-TEK
innovations in nucleic acid isolation

Product Manual

E.Z.N.A.® MicroElute Genomic DNA Kit

D3096-00	5 preps
D3096-01	50 preps
D3096-02	200 preps

Manual Date: September 2022
Revision Number: v9.0

For Research Use Only

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E.Z.N.A.® MicroElute Genomic DNA Kit

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Introduction

The E.Z.N.A.® MicroElute Genomic DNA Kit provides a rapid and easy method for the isolation of genomic and mitochondrial DNA from small sample sizes or from a large volume of samples for consistent PCR and other downstream applications. This kit can be used for the isolation of genomic DNA from micro-dissected tissue, cultured cells, blood, dry blood, swabs, buffy coat, serum, urine and plasma. The kit allows single or multiple processing of samples. There is no need for phenol/chloroform extractions and time-consuming steps such as precipitation with isopropanol or ethanol are eliminated.

The E.Z.N.A.® MicroElute Genomic DNA Kit uses the reversible binding properties of the HiBind® matrix, a silica-based material, in combination with the MicroElute spin column technology to allow elution volume as small as 10 µL. A specially formulated buffer system allows genomic DNA up to 40 kb to bind to the matrix. Samples are lysed under denaturing conditions and then transferred to the MicroElute DNA Mini Columns to bind DNA. Cellular debris, hemoglobin, and other proteins are efficiently eliminated via three quick wash steps. High-quality DNA is eluted in sterile deionized water or low salt buffer.

New in this Edition:

September 2022
- Column equilibration protocol is now required for more consistent results.

May 2019
- Linear Acrylamide has been renamed LPA Buffer. This is a name change only. The formulation has not changed.

March 2019
- OB Protease Solution has been renamed Proteinase K Solution. This is a name change only. The formulation has not changed.

Kit Contents

Product	D3096-00	D3096-01	D3096-02
Purifications	5	50	200
MicroElute LE DNA Columns	5	50	200
2 mL Collection Tubes	10	100	400
BL Buffer	5 mL	35 mL	130 mL
TL Buffer	5 mL	35 mL	125 mL
HBC Buffer	5 mL	25 mL	80 mL
Proteinase K Solution	150 µL	1.2 mL	4.5 mL
LPA Buffer	25 µL	125 µL	500 µL
DNA Wash Buffer	2.5 mL	25 mL	3 x 25 mL
Elution Buffer	15 mL	30 mL	30 mL
User Manual	✓	✓	✓

Storage and Stability

All of the E.Z.N.A.® MicroElute Genomic DNA Kit components are guaranteed for at least 12 months from the date of purchase when stored as follows. Proteinase K Solution can be stored at room temperature for up to 12 months. For long-term storage, store Proteinase K Solution at 2-8°C. During shipment or storage in cool ambient conditions, precipitates may form in BL Buffer. Dissolve such deposits by warming the solution at 37°C and gently shaking.

Determination of DNA Quality and Quantity

The total DNA yield can be determined by a spectrophotometer using deionized water, Tris-HCl buffer, or Elution Buffer as blank. DNA concentration is calculated as:

[DNA] = (Absorbance260) x (0.05 µg/ µL) x (Dilution factor)

The quality of DNA can be assessed by measuring absorbance at both 260 nm and at 280 nm. A ratio of (A_{260}/A_{280}) of 1.7-1.9 corresponds to 85%-95% purity. Yields vary with both amount and type of sample used.

Preparing Reagents

1. Dilute DNA Wash Buffer with 100% ethanol as follows and store at room temperature.

Kit	100% Ethanol to be Added
D3096-00	10 mL
D3096-01	100 mL
D3096-02	100 mL per bottle

2. Dilute HBC Buffer with 100% isopropanol as follows and store at room temperature.

Kit	100% Isopropanol to be Added
D3096-00	2 mL
D3096-01	10 mL
D3096-02	32 mL

E.Z.N.A.® MicroElute Genomic DNA Kit - Protocol for Tissue Samples

The following protocol allows for isolation of genomic DNA from up to 10 mg tissue. Yields will vary depending on source. To purify very small amounts of DNA from a sample, such as low volumes of blood (<10 µL) or micro-dissected tissues, we recommend adding LPA Buffer to BL Buffer to enhance the DNA binding ability of the column. In most cases, adding 5-10 µg (1-2 µL) per sample is sufficient.

Materials and Equipment to be Supplied by User:

- Microcentrifuge capable of at least 13,000*g*
- Water baths, heat blocks, or incubators capable of 55°C and 70°C
- Shaking water bath capable of 55°C
- Vortexer
- Nuclease-free 1.5 mL or 2 mL microcentrifuge tubes
- 100% ethanol
- 100% isopropanol
- 3M NaOH
- Sterile deionized water
- Optional: RNase A (25 mg/mL)

Before Starting:

- Prepare DNA Wash Buffer and HBC Buffer according to the "Preparing Reagents" section on Page 4
- Set water baths, heat blocks, or incubators to 55°C and 70°C
- Heat Elution Buffer to 70°C

1. Mince up to 10 mg tissue and transfer to a 1.5 mL microcentrifuge tube (not provided).

2. Add 200 µL TL Buffer

3. Add 20 µL Proteinase K Solution. Vortex to mix thoroughly.

4. Incubate at 55°C in a shaking water bath.

   Note: If a shaking water bath is not available, vortex the sample every 10-20 minutes. Lysis time will depend on the amount and type of tissue, but is usually less than 3 hours. Lysis can proceed overnight.

5. Centrifuge at maximum speed (13,000*g*) for 2 minutes.

6. Transfer the supernatant to a clean 1.5 mL microcentrifuge tube (not provided).

Optional: Certain tissues such as liver have high levels of RNA which will be co-purified with DNA using this kit. While it will not interfere with PCR, the RNA may be removed at this point. Add 5 µL RNase A (25 mg/mL, assuming a sample size of 10 mg) and incubate at room temperature for 2 minutes. Proceed to Step 7.

7. Add 220 µL BL Buffer. Vortex to mix thoroughly.

   Note: If LPA Buffer is needed, add 1-2 µL LPA Buffer to 220 µL BL Buffer.

8. Incubate at 70°C for 10 minutes.

9. Add 220 µL 100% ethanol. Vortex at maximum speed for 15 seconds. A precipitate may form at this point; it will not interfere with DNA isolation.

10. Briefly centrifuge to bring down any liquid from the top of the lid.

11. Insert a MicroElute® LE DNA Column into a 2 mL Collection Tube (provided) and follow the column equilibration steps listed below:

Protocol for Column Equilibration:

1. Add 100 µL 3M NaOH to the MicroElute® LE DNA Column.
2. Centrifuge at 10,000*g* for 30 seconds.
3. Add 500 µL sterile deionized water to the MicroElute® LE DNA Column.
4. Centrifuge at 10,000*g* for 30 seconds.
5. Discard the filtrate and reuse the collection tube.

12. Transfer the sample from Step 10 (including any precipitate that may have formed) to the MicroElute LE DNA Column.

13. Centrifuge at maximum speed for 1 minute.

14. Discard the filtrate and the Collection Tube.

15. Transfer the MicroElute LE DNA Column to a new 2 mL Collection Tube.

16. Add 500 µL HBC Buffer.

    Note: HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 4 for instructions.

17. Centrifuge at maximum speed for 30 seconds.

18. Discard the filtrate and reuse the Collection Tube.

19. Add 700 µL DNA Wash Buffer.

    Note: DNA Wash Buffer must be diluted with 100% ethanol prior to use. Please see the Preparing Reagents section on Page 4 for instructions.

20. Centrifuge at maximum speed for 1 minute.

21. Discard the filtrate and reuse the Collection Tube.

22. Repeat Steps 19-21 for a second DNA Wash Buffer wash step.

23. Centrifuge the empty MicroElute LE DNA Column at maximum speed for 2 minutes to dry the membrane.

    Note: It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

24. Transfer the MicroElute LE DNA Column to a nuclease-free 1.5 or 2 mL microcentrifuge tube (not provided).

25. Add 10-50 µL Elution Buffer (or sterile deionized water) heated to 70°C.

    Note: Smaller elution volumes will increase DNA concentration but decrease yield. Elution volumes greater than 200 µL are not recommended.

26. Let sit at room temperature for 3 minutes.

27. Centrifuge at maximum speed for 1 minute.

28. Store DNA at -20°C.

E.Z.N.A.® MicroElute Genomic DNA Kit - Protocol for Small Volumes of Blood, Serum, or Body Fluids

This protocol is designed for the rapid isolation of DNA from 1-100 µL blood (treated with EDTA, citrate, or heparin-based anticoagulants), serum, plasma, buffy coat, saliva, and urine. To purify very small amounts of DNA from a sample, such as low volumes of blood (<10 µL) or micro-dissected tissues, we recommend adding LPA Buffer to BL Buffer to enhance the DNA binding ability of the column. In most cases, adding 5-10 µg (1-2 µL) per sample is sufficient.

Materials and Equipment to be Supplied by User:

- Microcentrifuge capable of at least 13,000*g*
- Water bath, heat block, or incubator capable of 70°C
- Vortexer
- Nuclease-free 1.5 mL or 2 mL microcentrifuge tubes
- 100% isopropanol
- 100% ethanol
- PBS
- 3M NaOH
- Sterile deionized water

Before Starting:

- Prepare DNA Wash Buffer and HBC Buffer according to the "Preparing Reagents" section on Page 4
- Set water bath, heat block, or incubator to 70°C
- Heat Elution Buffer to 70°C

1. Add 1-100 µL sample (the sample must be at room temperature) to a 1.5 mL microcentrifuge tube (not provided).

2. Adjust the sample volume to 100 µL with PBS.

3. Add 20 µL Proteinase K Solution. Vortex to mix thoroughly.

4. Add 120 µL BL Buffer. Vortex to mix thoroughly.

5. Incubate at 70°C for 10 minutes.

6. Add 160 µL isopropanol. Vortex at maximum speed for 15 seconds. A precipitate may form at this point; it will not interfere with DNA isolation.

7. Briefly centrifuge to bring down any liquid from the top of the lid.

8. Insert a MicroElute® LE DNA Column into a 2 mL Collection Tube (provided) and follow the column equilibration steps listed below:

Protocol for Column Equilibration:

1. Add 100 µL 3M NaOH to the MicroElute® LE DNA Column.
2. Centrifuge at 10,000*g* for 30 seconds.
3. Add 500 µL sterile deionized water to the MicroElute® LE DNA Column.
4. Centrifuge at 10,000*g* for 30 seconds.
5. Discard the filtrate and reuse the collection tube.

9. Transfer the sample from Step 7 (including any precipitate that may have formed) to the MicroElute LE DNA Column.

10. Centrifuge at maximum speed for 1 minute.

11. Discard the filtrate and the Collection Tube.

12. Transfer the MicroElute LE DNA Column to a new 2 mL Collection Tube.

13. Add 500 µL HBC Buffer.

    Note: HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 4 for instructions.

14. Centrifuge at maximum speed for 30 seconds.

15. Discard the filtrate and reuse the Collection Tube.

16. Add 700 µL DNA Wash Buffer.

    Note: DNA Wash Buffer must be diluted with 100% ethanol prior to use. Please see the Preparing Reagents section on Page 4 for instructions.

17. Centrifuge at maximum speed for 1 minute.

18. Discard the filtrate and reuse the Collection Tube.

19. Repeat Steps 16-18 for a second DNA Wash Buffer wash step.

20. Centrifuge the empty MicroElute LE DNA Column at maximum speed for 2 minutes to dry the membrane.

    Note: It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

21. Transfer the MicroElute LE DNA Column to a nuclease-free 1.5 or 2 mL microcentrifuge tube (not provided).

22. Add 10-50 µL Elution Buffer (or sterile deionized water) heated to 70°C.

    Note: Smaller elution volumes will increase DNA concentration but decrease yield. Elution volumes greater than 200 µL are not recommended.

23. Let sit at room temperature for 3 minutes.

24. Centrifuge at maximum speed for 1 minute.

25. Store DNA at -20°C.

E.Z.N.A.® MicroElute Genomic DNA Kit - Protocol for Dried Blood, Body Fluids, and Semen Spots

Dried blood, body fluids, and semen samples on filter paper can be processed using the following method.

Materials and Equipment to be Supplied by User:

- Microcentrifuge capable of at least 13,000*g*
- Water baths, heat blocks, or incubators capable of 55°C and 70°C
- Shaking water bath capable of 55°C
- Vortexer
- Nuclease-free 1.5 mL or 2 mL microcentrifuge tubes
- 100% ethanol
- 100% isopropanol
- 3M NaOH
- Sterile deionized water
- Optional: Dithiothreitol (DTT)

Before Starting:

- Prepare DNA Wash Buffer and HBC Buffer according to the "Preparing Reagents" section on Page 4
- Set water baths, heat blocks, or incubators to 55°C and 70°C
- Heat Elution Buffer to 70°C

1. Cut or punch out the blood (or other sample) spot from the filter paper. Tear or cut the filter paper into small pieces and place into a 1.5 mL microcentrifuge tube (not provided).

   Note: Use 1-3 punched circles (~3 mm diameter) for each DNA isolation.

2. Add 200 µL TL Buffer and 20 µL Proteinase K Solution. If you are processing semen spots, add 20 µL DTT to each sample.

3. Incubate at 55°C for 45-60 minutes in a shaking water bath.

   Note: If a shaking water bath is not available, vortex the sample every 10-20 minutes.

4. Briefly centrifuge to bring down any liquid from the top of the tube.

5. Add 220 µL BL Buffer. Vortex to mix thoroughly.

   Note: If only one punch card is processed, add 1 µl LPA Buffer to the sample.

6. Incubate at 70°C for 10 minutes. Vortex at maximum speed for 10 seconds several times during incubation.

7. Centrifuge at maximum speed for 5 minutes.

8. Transfer the cleared lysate to a new 1.5 mL microcentrifuge tube (not provided).

   Note: For maximum yield collect any remaining liquid from the paper and transfer the entire sample, including paper, into an Omega Homogenizer Column (not provided, Cat# HCR003) and centrifuge at maximum speed for 2 minutes to collect all of the lysate.

9. Add 220 µL 100% ethanol. Vortex at maximum speed for 15 seconds.

10. Briefly centrifuge to bring down any liquid from the top of the lid.

11. Insert a MicroElute® LE DNA Column into a 2 mL Collection Tube (provided) and follow the column equilibration steps listed below:

Protocol for Column Equilibration:

1. Add 100 µL 3M NaOH to the MicroElute® LE DNA Column.
2. Centrifuge at 10,000*g* for 30 seconds.
3. Add 500 µL sterile deionized water to the MicroElute® LE DNA Column.
4. Centrifuge at 10,000*g* for 30 seconds.
5. Discard the filtrate and reuse the collection tube.

12. Transfer the sample from Step 10 (including any precipitate that may have formed) to the MicroElute LE DNA Column.

13. Centrifuge at maximum speed for 1 minute.

14. Discard the filtrate and the Collection Tube.

15. Transfer the MicroElute LE DNA Column to a new 2 mL Collection Tube.

16. Add 500 µL HBC Buffer.

    Note: HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 4 for instructions.

17. Centrifuge at maximum speed for 30 seconds.

18. Discard the filtrate and reuse the Collection Tube.

19. Add 700 µL DNA Wash Buffer.

    Note: DNA Wash Buffer must be diluted with 100% ethanol prior to use. Please see the Preparing Reagents section on Page 4 for instructions.

20. Centrifuge at maximum speed for 1 minute.

21. Discard the filtrate and reuse the Collection Tube.

22. Repeat Steps 19-21 for a second DNA Wash Buffer wash step.

23. Centrifuge the empty MicroElute LE DNA Column at maximum speed for 2 minutes to dry the membrane.

    Note: It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

24. Transfer the MicroElute LE DNA Column to a nuclease-free 1.5 or 2 mL microcentrifuge tube (not provided).

25. Add 10-50 µL Elution Buffer (or sterile deionized water) heated to 70°C.

    Note: Smaller elution volumes will increase DNA concentration but decrease yield. Elution volumes greater than 200 µL are not recommended.

26. Let sit at room temperature for 3 minutes.

27. Centrifuge at maximum speed for 1 minute.

28. Store DNA at -20°C.

E.Z.N.A.® MicroElute Genomic DNA Kit - Protocol for Swabs

This protocol is designed for the isolation of genomic DNA from semen swabs, blood swabs, and buccal swabs.

Materials and Equipment to be Supplied by User:

- Microcentrifuge capable of at least 13,000*g*
- Water baths, heat blocks, or incubators capable of 55°C and 70°C
- Shaking water bath capable of 55°C
- Vortexer
- Nuclease-free 2 mL microcentrifuge tubes
- 100% ethanol
- 100% isopropanol
- 3M NaOH
- Sterile deionized water
- Optional: Dithiothreitol (DTT)

Before Starting:

- Prepare DNA Wash Buffer and HBC Buffer according to the "Preparing Reagents" section on Page 4
- Set water baths, heat blocks, or incubators to 55°C and 70°C
- Heat Elution Buffer to 70°C

1. Place the swab in a 2 mL microcentrifuge tube.

2. Add 600 µL TL Buffer and 20 µL Proteinase K Solution. Vortex at maximum speed for 30 seconds. If you are processing semen swabs, add 20 µL DTT to each sample.

3. Incubate at 55°C for 60 minutes in a shaking water bath.

   Note: If a shaking water bath is not available, vortex the sample every 10-20 minutes.

4. Briefly centrifuge to bring down any liquid from the top of the tube.

5. Add 620 µL BL Buffer. Vortex at maximum speed for 20 seconds.

6. Incubate at 70°C for 10 minutes. Vortex at maximum speed for 10 seconds several times during incubation.

7. Briefly centrifuge to bring down any liquid from the top of the tube

8. Transfer the cleared lysate to a new 1.5 mL microcentrifuge tube (not provided).

9. Add 620 µL 100% ethanol. Vortex at maximum speed for 15 seconds.

10. Briefly centrifuge to bring down any liquid from the top of the lid.

11. Insert a MicroElute® LE DNA Column into a 2 mL Collection Tube (provided) and follow the column equilibration steps listed below:

Protocol for Column Equilibration:

1. Add 100 µL 3M NaOH to the MicroElute® LE DNA Column.
2. Centrifuge at 10,000*g* for 30 seconds.
3. Add 500 µL sterile deionized water to the MicroElute® LE DNA Column.
4. Centrifuge at 10,000*g* for 30 seconds.
5. Discard the filtrate and reuse the collection tube.

12. Transfer the sample from Step 10 (including any precipitate that may have formed) to the MicroElute LE DNA Column.

13. Centrifuge at maximum speed for 1 minute.

14. Repeat Steps 12-13 until all the sample has been transferred to the MicroElute LE DNA Column.

15. Discard the filtrate and the Collection Tube.

16. Transfer the MicroElute LE DNA Column to a new 2 mL Collection Tube.

17. Add 500 µL HBC Buffer.

    Note: HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 4 for instructions.

18. Centrifuge at maximum speed for 30 seconds.

19. Discard the filtrate and reuse the Collection Tube.

20. Add 700 µL DNA Wash Buffer.

    Note: DNA Wash Buffer must be diluted with 100% ethanol prior to use. Please see the Preparing Reagents section on Page 4 for instructions.

21. Centrifuge at maximum speed for 1 minute.

22. Discard the filtrate and reuse the Collection Tube.

23. Repeat Steps 20-22 for a second DNA Wash Buffer wash step.

24. Centrifuge the empty MicroElute LE DNA Column at maximum speed for 2 minutes to dry the membrane.

    Note: It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

25. Transfer the MicroElute LE DNA Column to a nuclease-free 2 mL microcentrifuge tube (not provided).

26. Add 10-50 µL Elution Buffer (or sterile deionized water) heated to 70°C.

    Note: Smaller elution volumes will increase DNA concentration but decrease yield. Elution volumes greater than 200 µL are not recommended.

27. Let sit at room temperature for 3 minutes.

28. Centrifuge at maximum speed for 1 minute.

29. Store DNA at -20°C.

E.Z.N.A.® MicroElute Genomic DNA Kit - Protocol for Forensic Samples

This protocol is designed for isolation of genomic DNA from forensic samples such as hair, cigarette butts, nail clippings, material stained with blood, saliva, or semen.

Materials and Equipment to be Supplied by User:

- Microcentrifuge capable of at least 13,000*g*
- Water baths capable of 55°C and 70°C
- Shaking water bath capable of 55°C
- Vortexer
- Nuclease-free 1.5 mL or 2 mL microcentrifuge tubes
- 100% ethanol
- 100% isopropanol
- 3M NaOH
- Sterile deionized water
- Optional: Dithiothreitol (DTT)

Before Starting:

- Prepare DNA Wash Buffer and HBC Buffer according to the "Preparing Reagents" section on Page 4
- Set water baths to 55°C and 70°C
- Heat Elution Buffer to 70°C

1. Cut the sample to small pieces and place into a 2 mL microcentrifuge tube (not provided).

2. Add 300 µL TL Buffer. Vortex to mix thoroughly. If you are processing semen stains, add 20 µL DTT to each sample.

3. Add 20 µL Proteinase K Solution. Vortex at maximum speed for 30 seconds.

4. Incubate mixture at 55°C for 60 minutes in a shaking water bath.

   Note: If a shaking water bath is not available, vortex the sample every 10-20 minutes. Lysis time will depend on the amount and type of sample but is approximately one hour. Extend incubation time of hair samples until lysis is complete. Lysis can proceed overnight for larger samples of nail clippings.

5. Briefly centrifuge to bring down any liquid from the top of the tube.

6. Add 320 µL BL Buffer. Vortex at maximum speed for 20 seconds.

7. Incubate at 70°C for 10 minutes. Vortex at maximum speed for 10 seconds several times during incubation.

8. Centrifuge at maximum speed for 5 minutes.

9. Transfer the cleared supernatant to a new 2 mL microcentrifuge tube (not provided).

10. Add 320 µL 100% ethanol. Vortex at maximum speed for 15 seconds.

11. Briefly centrifuge to bring down any liquid from the top of the lid.

12. Insert a MicroElute® LE DNA Column into a 2 mL Collection Tube (provided) and follow the column equilibration steps listed below:

Protocol for Column Equilibration:

1. Add 100 µL 3M NaOH to the MicroElute® LE DNA Column.
2. Centrifuge at 10,000*g* for 30 seconds.
3. Add 500 µL sterile deionized water to the MicroElute® LE DNA Column.
4. Centrifuge at 10,000*g* for 30 seconds.
5. Discard the filtrate and reuse the collection tube.

13. Transfer the sample from Step 11 (including any precipitate that may have formed) to the MicroElute LE DNA Column.

14. Centrifuge at maximum speed for 1 minute.

15. Repeat Steps 13-14 until all the sample has been transferred to the MicroElute LE DNA Column.

16. Discard the filtrate and the Collection Tube.

17. Transfer the MicroElute LE DNA Column to a new 2 mL Collection Tube.

18. Add 500 µL HBC Buffer.

    Note: HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 4 for instructions.

19. Centrifuge at maximum speed for 30 seconds.

20. Discard the filtrate and reuse the Collection Tube.

21. Add 700 µL DNA Wash Buffer.

    Note: DNA Wash Buffer must be diluted with 100% ethanol prior to use. Please see the Preparing Reagents section on Page 4 for instructions.

22. Centrifuge at maximum speed for 1 minute.

23. Discard the filtrate and reuse the Collection Tube.

24. Repeat Steps 21-23 for a second DNA Wash Buffer wash step.

25. Centrifuge the empty MicroElute LE DNA Column at maximum speed for 2 minutes to dry the membrane.

    Note: It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

26. Transfer the MicroElute LE DNA Column to a nuclease-free 1.5 or 2 mL microcentrifuge tube (not provided).

27. Add 10-50 µL Elution Buffer (or sterile deionized water) heated to 70°C.

    Note: Smaller elution volumes will increase DNA concentration but decrease yield. Elution volumes greater than 200 µL are not recommended.

28. Let sit at room temperature for 3 minutes.

29. Centrifuge at maximum speed for 1 minute.

30. Store DNA at -20°C.

Troubleshooting Guide

Please use this guide to troubleshoot any problems that may arise. For further assistance, please contact the technical support staff , toll free, **at 1-800-832-8896.**

Problem	Cause	Solution
Clogged Column	Incomplete lysis	Extend incubation time of lysis with TL Buffer and protease. Add the correct volume of BL Buffer and incubate for specified time at 70ºC. It may be necessary to extend incubation time by 10 minutes.
	Sample too large	Do not use greater than the recommended amount of starting material. For larger samples, divide into multiple tubes.
	Incomplete homogenization	Pulverize material as indicated in liquid nitrogen to obtain a fine powder.
Low or No DNA yield	Clogged column	See above
	Poor elution	Repeat elution or increase elution volume. Incubate the column at 70°C for 5 minutes before centrifugation.
	Poor binding to column	Follow the protocol closely when adjusting the binding conditions.
	Improper washing	DNA Wash Buffer must be diluted with 100% ethanol before use.
		HBC Buffer must be diluted with 100% isopropanol before use.
	Poor cell lysis	Increase incubation time with TL Buffer. An overnight incubation may be necessary.
	Incomplete homogenization	Pulverize starting material as indicated in liquid nitrogen to obtain a fine powder.
	100% ethanol was not added before adding sample to column	Before applying DNA sample to column, add TL Buffer and 100% ethanol.
	Ethanol was not added to the DNA Wash Buffer	Dilute DNA Wash Buffer with the indicated volume of 100% ethanol before use.
	Column Equilibration not performed	Perform the column equilibration protocol as instructed in the manual for consistent results.

For more purification solutions, visit **www.omegabiotek.com**

AVAILABLE FORMATS


Spin Columns

96-Well Silica Plates


Mag Beads

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Plasmid


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FFPE


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