



## E.Z.N.A.<sup>®</sup> Blood DNA Mini Kit

D3392-02	200 preps
D3392-03	600 preps

**Manual Date: November 2025**  
**Revision Number: v9.0**

**For Research Use Only**



# E.Z.N.A.® Blood DNA Mini Kit

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# Introduction

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The E.Z.N.A.® family of products is an innovative system that radically simplifies the extraction and purification of nucleic acids from a variety of sources. The key to this system is the new HiBind® matrix that specifically, but reversibly, binds DNA or RNA under optimal conditions allowing proteins and other contaminants to be removed. Nucleic acids are easily eluted with deionized water or a low salt buffer.

The E.Z.N.A.® Blood DNA Mini Kit provides an easy and rapid method for the isolation of genomic DNA for consistent PCR and Southern analysis. Up to 250 µL fresh, frozen, or anticoagulated whole blood can be readily processed at one time. The E.Z.N.A.® Blood DNA Mini Kit can also be used for the preparation of genomic DNA from buffy coat, serum, plasma, saliva, buccal swabs, and other body fluids. The E.Z.N.A.® Blood DNA Kit allows for single or multiple simultaneous processing of multiple samples. There is no need for phenol/chloroform extractions, and time-consuming steps are eliminated (e.g. precipitation using isopropanol or ethanol). Purified DNA obtained with the E.Z.N.A.® Blood DNA Kit is ready for applications such as PCR, restriction digestion, and Southern blotting.

## Benefits of the E.Z.N.A.® Blood DNA Mini Kit

- Optimized buffers that guarantee pure DNA
- No organic extractions
- Purified DNA can be directly used in most downstream applications

## Binding Capacity

Each HiBind® DNA Mini Column can bind approximately 100 µg DNA. Using greater than 250 µL whole blood or buffy coat is not recommended.

## New in this Edition:

November 2025

- Addition of Fresh or Frozen Sperm Protocol.

April 2024

- Addition of Warnings and Safety Information.

June 2023:

- General revision.

March 2022:

- A new 600 prep kit (D3392-03) has been added and is now available for purchase.

# Illustrated Protocol

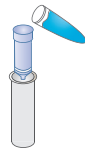
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Lyse



Adjust Binding  
Conditions



Bind



Wash 3X



Dry



Elute

# Kit Contents

Product	D3392-02	D3392-03
Purifications	200 preps	600 preps
HiBind® DNA Mini Columns	200	600
2 mL Collection Tubes	400	1200
BL Buffer	60 mL	3 x 110 mL
HBC Buffer	80 mL	250 mL
DNA Wash Buffer	3 x 25 mL	200 mL
Elution Buffer	250 mL	300 mL
Proteinase K Solution	6 mL	18 mL
User Manual	✓	✓

## Storage and Stability

All of the E.Z.N.A.® Blood DNA Mini Kit components are guaranteed for 12 months from the date of purchase when stored as follows. Proteinase K Solution can be stored at room temperature for 12 months. For long-term storage (>12 months), store Proteinase K Solution at 2-8°C. Store all other components at room temperature and away from bright light. During shipment or storage under cool ambient conditions, a precipitate may form in some buffers. Dissolve such deposits by warming the solution to 37°C and gently shaking.

# Warnings and Safety Information

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## Warnings

This kit is for research use only.

Please read all instructions carefully before using the kit.

Decontaminate and dispose of all potentially infectious materials in accordance with applicable local, state, and national regulations. Please refer to safety data sheets (SDSs) for information on disposal of different components included in this kit.

## Safety Information

All chemicals and biological materials are potentially hazardous. Biological samples such as plasma, serum, tissues, body fluids, blood etc. are potentially infectious and must be treated as biohazardous materials. Conduct all work in properly equipped facilities following universal precautions and using appropriate personal safety equipment such as disposable gloves, lab coats, safety glasses etc. as required by policies and procedures outlined by your facility. Please refer to safety data sheets (SDSs) for information on safe handling, transport and disposal of different components included in this kit. SDSs are made available in PDF format on the product page at [www.omegabiotek.com](http://www.omegabiotek.com). Discard all waste in accordance with the local safety regulations.

Some of the buffers included in the product contain guanidine-based chaotropic agents, which can form highly reactive compounds when combined with bleach. DO NOT add bleach or acidic solutions to guanidine-containing waste. Please access the SDSs online for detailed information on the reagents.

## Preparing Reagents

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1. Dilute HBC Buffer with 100% isopropanol as follows and store at room temperature.

<b>Kit</b>	<b>100% Isopropanol to be Added</b>
D3392-02	32 mL
D3392-03	100 mL

2. Dilute DNA Wash Buffer with 100% ethanol as follows and store at room temperature.

<b>Kit</b>	<b>100% Ethanol to be Added</b>
D3392-02	100 mL per bottle
D3392-03	800 mL

# E.Z.N.A.® Blood DNA Mini Kit

## Blood and Body Fluids Protocol

The procedure below has been optimized for the use with fresh or frozen blood samples up to 250  $\mu$ L in volume. Anti-coagulated blood, saliva, serum, buffy coat, or other body fluids can also be used. In addition,  $\leq 10^7$  of leukocytes or cultured cells may be used with this procedure. For DNA extraction from tissue and mouse tail we suggest that you use the E.Z.N.A.® Tissue DNA Kit (Product No. D3396). To isolate viral RNA from serum or other non-cellular body fluids the E.Z.N.A.® Viral RNA Kit (Product No. R6874) is recommended.

### Materials and Equipment to be Supplied by User:

- Tabletop microcentrifuge capable of at least 13,000g
- Water bath, incubator, or heat block capable of 65°C
- Vortexer
- Nuclease-free 2 mL microcentrifuge tubes
- 100% ethanol
- 100% isopropanol
- Optional: 10mM Tris-HCl or PBS
- Optional: RNase stock solution (50 mg/mL; when RNA-free genomic DNA is required)

### Before Starting:

- Prepare HBC Buffer and DNA Wash Buffer according to the directions in the "Preparing Reagents" section on Page 6
  - Set water bath, incubator, or heat block to 65°C
  - Heat the Elution Buffer to 65°C
1. Transfer the sample into a nuclease-free 2 mL microcentrifuge tube (not provided) and bring the volume up to 250  $\mu$ L with 10mM Tris-HCl (not provided), PBS (not provided), or Elution Buffer.
  2. Add 25  $\mu$ L Proteinase K Solution and 250  $\mu$ L BL Buffer. Vortex at maximum speed for 15 seconds.  
  
**Optional:** If RNA-free genomic DNA is required, add 5  $\mu$ L RNase A (50 mg/mL).
  3. Incubate for 10 minutes at 65°C. Vortex briefly once during incubation.
  4. Add 260  $\mu$ L 100% ethanol (not provided). Vortex at maximum speed for 20 seconds.

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5. Centrifuge briefly to collect any drops from the inside of the lid.
6. Insert a HiBind<sup>®</sup> DNA Mini Column into a 2 mL collection tube.
7. Transfer the entire sample to the column.
8. Centrifuge at  $\geq 10,000g$  for 1 minute.
9. Discard the filtrate and the collection tube.
10. Insert the HiBind<sup>®</sup> DNA Mini Column into a new 2 mL collection tube.
11. Add 500  $\mu$ L HBC Buffer.  
**Note:** HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 6 for instructions.
12. Centrifuge at  $\geq 10,000g$  for 1 minute.
13. Discard the filtrate and reuse the collection tube.
14. Add 700  $\mu$ L DNA Wash Buffer.  
**Note:** DNA Wash Buffer must be diluted with 100% ethanol before use. Please see Page 6 for instructions.
15. Centrifuge at 10,000g for 1 minute.
16. Discard the filtrate and reuse the collection tube.

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17. Repeat Steps 14-16 for a second DNA Wash Buffer step.
18. Centrifuge the empty HiBind<sup>®</sup> DNA Mini Column at maximum speed ( $\geq 10,000g$ ) for 2 minutes to dry the column matrix.

**Note:** It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

19. Transfer the HiBind<sup>®</sup> DNA Mini Column into a nuclease-free 2 mL microcentrifuge tube.

20. Add 100-200  $\mu$ L Elution Buffer heated to 65°C.

21. Let sit for 5 minutes at room temperature.

**Note:** Incubating the HiBind<sup>®</sup> DNA Mini Column at 65°C rather than room temperature will give a modest increase in DNA yield per elution.

22. Centrifuge at  $\geq 13,000g$  for 1 minute.

23. Repeat Steps 20-22 for a second elution step.

**Note:** Any combination of the following steps can be used to help increase DNA yield.

- After adding the Elution Buffer, incubate the column for 5 minutes.
- Increase the elution volume.
- Repeat the elution step with fresh Elution Buffer (this may increase the yield, but decrease the concentration).
- Repeat the elution step using the eluate from the first elution (this may increase yield while maintaining elution volume).

24. Store eluted DNA at -20°C.

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## Buccal Swabs Protocol

### **IMPORTANT:** Additional Reagents May Be Required

This protocol requires more reagent volumes exceeding those provided in the standard kit protocol, depending on the number of samples processed. The following reagents will need to be purchased separately:

- [BL Buffer](#)

### **Materials and Equipment to be Supplied by User:**

- Tabletop microcentrifuge capable of at least 13,000g
- Water bath, incubator, or heat block capable of 65°C
- Vortexer
- Nuclease-free 2 mL microcentrifuge tubes
- 100% ethanol
- 100% isopropanol
- PBS
- Optional: RNase stock solution (50 mg/mL; when RNA-free genomic DNA is required)

### **Before Starting:**

- Prepare HBC Buffer and DNA Wash Buffer according to the directions in the “Preparing Reagents” section on Page 6
- Set water bath, incubator, or heat block to 65°C
- Heat the Elution Buffer to 65°C

1. Place the buccal swab in a nuclease-free 2 mL microcentrifuge tube (not provided).
2. Add 500  $\mu$ L PBS (not provided).

**Optional:** If RNA-free genomic DNA is required, add 5  $\mu$ L RNase A (50 mg/mL).

3. Add 25  $\mu$ L Proteinase K Solution and 500  $\mu$ L BL Buffer. Vortex at maximum speed for 30 seconds.
4. Incubate for 10 minutes at 65°C.
5. Discard the buccal swab.

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6. Add 500  $\mu$ L 100% ethanol (not provided). Vortex at maximum speed for 20 seconds.
7. Centrifuge briefly to collect any drops from the inside of the lid.
8. Insert a HiBind<sup>®</sup> DNA Mini Column into a 2 mL collection tube.
9. Transfer 750  $\mu$ L sample to the column.
10. Centrifuge at 10,000*g* for 1 minute.
11. Discard the filtrate and reuse the collection tube.
12. Repeat Steps 9-11 until all the sample has been transferred to the column.
13. Place the HiBind DNA Mini Column into a new 2 mL collection tube.
14. Add 500  $\mu$ L HBC Buffer.  
**Note:** HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 6 for instructions.
15. Centrifuge at  $\geq$ 10,000*g* for 1 minute.
16. Discard the filtrate and reuse collection tube.
17. Add 700  $\mu$ L DNA Wash Buffer.  
**Note:** DNA Wash Buffer must be diluted with 100% ethanol before use. Please see Page 6 for instructions.
18. Centrifuge at 10,000*g* for 1 minute.

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19. Discard the filtrate and reuse the collection tube.
20. Repeat Steps 17-19 for a second DNA Wash Buffer step.
21. Centrifuge the empty HiBind<sup>®</sup> DNA Mini Column at maximum speed ( $\geq 10,000g$ ) for 2 minutes to dry the column matrix.

**Note:** It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

22. Transfer the HiBind<sup>®</sup> DNA Mini Column into a nuclease-free 2 mL microcentrifuge tube.
23. Add 100-200  $\mu$ L Elution Buffer heated to 65°C.
24. Let sit for 5 minutes at room temperature.

**Note:** Incubating the HiBind<sup>®</sup> DNA Mini Column at 65°C rather than room temperature will give a modest increase in DNA yield per elution.

25. Centrifuge at  $\geq 13,000g$  for 1 minute.
26. Repeat Steps 23-25 for a second elution step.

**Note:** Any combination of the following steps can be used to help increase DNA yield.

- After adding the Elution Buffer, incubate the column for 5 minutes.
- Increase the elution volume.
- Repeat the elution step with fresh Elution Buffer (this may increase the yield, but decrease the concentration).
- Repeat the elution step using the eluate from the first elution (this may increase yield while maintaining elution volume).

27. Store eluted DNA at -20°C.

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## Dried Blood Protocol

### Materials and Equipment to be Supplied by User:

- Tabletop microcentrifuge capable of at least 13,000g
- Water bath, incubator, or heat block capable of 65°C
- Vortexer
- Nuclease-free 2 mL microcentrifuge tubes
- 100% ethanol
- 100% isopropanol
- PBS
- Optional: RNase stock solution (50 mg/mL; when RNA-free genomic DNA is required)

### Before Starting:

- Prepare HBC Buffer and DNA Wash Buffer according to the directions in the “Preparing Reagents” section on Page 6
  - Set water bath, incubator, or heat block to 65°C
  - Heat the Elution Buffer to 65°C
1. Cut or punch-out the blood spot from the filter paper (up to 200  $\mu$ L blood can be used per spot). Tear or cut the filter paper into small pieces and place them into a nuclease-free 2 mL microcentrifuge tube (not provided).
  2. Add 250  $\mu$ L PBS (not provided).
  3. Incubate for 1 hour at 65°C. Vortex briefly every 20 minutes.
  4. Add 25  $\mu$ L Proteinase K Solution. Vortex at maximum speed for 15 seconds.
  5. Incubate for 30 minutes at 65°C. Vortex briefly several times during incubation.
  6. Centrifuge at  $\geq 13,000g$  for 5 minutes.
  7. Transfer the supernatant to a nuclease-free 2 mL microcentrifuge tube.

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8. Add 1 volume BL Buffer and 1 volume 100% ethanol (not provided). Vortex to mix thoroughly.
9. Centrifuge briefly to collect any drops from the inside of the lid.
10. Insert a HiBind<sup>®</sup> DNA Mini Column into a 2 mL collection tube.
11. Transfer the entire sample to the column.
12. Centrifuge at  $\geq 10,000g$  for 1 minute.
13. Discard the filtrate and the collection tube.
14. Insert the HiBind<sup>®</sup> DNA Mini Column into a new 2 mL collection tube.
15. Add 500  $\mu$ L HBC Buffer.  
**Note:** HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 6 for instructions.
16. Centrifuge at  $\geq 10,000g$  for 1 minute.
17. Discard the filtrate and reuse the collection tube.
18. Add 700  $\mu$ L DNA Wash Buffer.  
**Note:** DNA Wash Buffer must be diluted with 100% ethanol before use. Please see Page 6 for instructions.
19. Centrifuge at 10,000g for 1 minute.
20. Discard the filtrate and reuse the collection tube.

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21. Repeat Steps 18-20 for a second DNA Wash Buffer step.
22. Centrifuge the empty HiBind<sup>®</sup> DNA Mini Column at maximum speed ( $\geq 10,000g$ ) for 2 minutes to dry the column matrix.

**Note:** It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

23. Transfer the HiBind<sup>®</sup> DNA Mini Column into a nuclease-free 2 mL microcentrifuge tube.

24. Add 100-200  $\mu\text{L}$  Elution Buffer heated to 65°C.

25. Let sit for 5 minutes at room temperature.

**Note:** Incubating the HiBind<sup>®</sup> DNA Mini Column at 65°C rather than room temperature will give a modest increase in DNA yield per elution.

26. Centrifuge at  $\geq 13,000g$  for 1 minute.

27. Repeat Steps 24-26 for a second elution step.

**Note:** Any combination of the following steps can be used to help increase DNA yield.

- After adding the Elution Buffer, incubate the column for 5 minutes.
- Increase the elution volume.
- Repeat the elution step with fresh Elution Buffer (this may increase the yield, but decrease the concentration).
- Repeat the elution step using the eluate from the first elution (this may increase yield while maintaining elution volume).

28. Store eluted DNA at -20°C.

**Note:** Blood spots from finger pricks usually contain no more than 50  $\mu\text{L}$  blood, and yield approximately 500 ng to 1  $\mu\text{g}$  DNA. This is usually sufficient for PCR analysis. To obtain higher DNA concentrations, elute with 50  $\mu\text{L}$  preheated elution buffer (volumes lower than 50  $\mu\text{L}$  greatly reduce yields). Alternatively, the first eluate can be used to perform a second elution.

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## Buffy Coat Protocol

The buffy coat fraction of whole blood is enriched with leukocytes and usually gives at least 5-fold more DNA than the same volume of blood. To prepare the buffy coat from fresh whole blood, simply centrifuge the sample at 3,000-4,000g for 10 minutes at room temperature. Three layers should form: a plasma upper layer, a buffy coat middle layer, and an erythrocyte bottom layer. Carefully aspirate the plasma, making sure not to disturb the layer of concentrated leukocytes. The buffy coat can be drawn off with a pipette and used directly in the E.Z.N.A.<sup>®</sup> Blood DNA Mini Kit or frozen at -70°C.

### **IMPORTANT:** Additional Reagents May Be Required

This protocol requires more reagent volumes exceeding those provided in the standard kit protocol, depending on the number of samples processed. The following reagents will need to be purchased separately:

- [BL Buffer](#)

### **Materials and Equipment to be Supplied by User:**

- Tabletop microcentrifuge capable of at least 13,000g
- Water bath, incubator, or heat block capable of 65°C
- Vortexer
- Nuclease-free 2 mL microcentrifuge tubes
- 100% ethanol
- 100% isopropanol
- Optional: 10mM Tris-HCl or PBS
- Optional: RNase stock solution (50 mg/mL; when RNA-free genomic DNA is required)

### **Before Starting:**

- Prepare HBC Buffer and DNA Wash Buffer according to the directions in the "Preparing Reagents" section on Page 6
  - Set water bath, incubator, or heat block to 65°C
  - Heat the Elution Buffer to 65°C
1. Transfer the sample into a nuclease-free 2 mL microcentrifuge tube (not provided) and bring the volume up to 500 µL with 10mM Tris-HCl (not provided), PBS (not provided), or Elution Buffer.

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2. Add 25  $\mu$ L Proteinase K Solution and 500  $\mu$ L BL Buffer. Vortex at maximum speed for 15 seconds.

**Optional:** If RNA-free genomic DNA is required, add 2  $\mu$ L RNase A (50 mg/mL).

3. Incubate for 10 minutes at 65°C. Vortex briefly once during incubation.
4. Add 500  $\mu$ L 100% ethanol (not provided). Vortex at maximum speed for 20 seconds.
5. Centrifuge briefly to collect any drops from the inside of the lid.
6. Insert a HiBind<sup>®</sup> DNA Mini Column into a 2 mL collection tube.
7. Transfer the 750  $\mu$ L sample to the column.
8. Centrifuge at  $\geq 10,000g$  for 1 minute.
9. Discard the filtrate and the collection tube.
10. Repeat Steps 7-9 until all the sample has been transferred to the column.
11. Insert the HiBind<sup>®</sup> DNA Mini Column into a new 2 mL collection tube.
12. Add 500  $\mu$ L HBC Buffer.

**Note:** HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 6 for instructions.

13. Centrifuge at  $\geq 10,000g$  for 1 minute.
14. Discard the filtrate and reuse the collection tube.

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15. Add 700  $\mu$ L DNA Wash Buffer.

**Note:** DNA Wash Buffer must be diluted with 100% ethanol before use. Please see Page 6 for instructions.

16. Centrifuge at 10,000g for 1 minute.

17. Discard the filtrate and reuse the collection tube.

18. Repeat Steps 15-17 for a second DNA Wash Buffer step.

19. Centrifuge the empty HiBind<sup>®</sup> DNA Mini Column at maximum speed ( $\geq 10,000g$ ) for 2 minutes to dry the column matrix.

**Note:** It is important to dry the column membrane before elution. Residual ethanol may interfere with downstream applications.

20. Transfer the HiBind<sup>®</sup> DNA Mini Column into a nuclease-free 2 mL microcentrifuge tube.

21. Add 100-200  $\mu$ L Elution Buffer heated to 65°C.

22. Let sit for 5 minutes at room temperature.

**Note:** Incubating the HiBind<sup>®</sup> DNA Mini Column at 65°C rather than room temperature will give a modest increase in DNA yield per elution.

23. Centrifuge at  $\geq 13,000g$  for 1 minute.

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24. Repeat Steps 21-23 for a second elution step.

**Note:** Any combination of the following steps can be used to help increase DNA yield.

- After adding the Elution Buffer, incubate the column for 5 minutes.
- Increase the elution volume.
- Repeat the elution step with fresh Elution Buffer (this may increase the yield, but decrease the concentration).
- Repeat the elution step using the eluate from the first elution (this may increase yield while maintaining elution volume).

25. Store eluted DNA at -20°C.

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## Fresh or Frozen Semen

This protocol can be used for fresh or frozen semen samples with equal efficiency. Frozen samples must be completely thawed before use. Note that lysis time will vary depending on the size and density of the source material.

### **IMPORTANT:** Additional Reagents May Be Required

This protocol requires more reagent volumes exceeding those provided in the standard kit protocol, depending on the number of samples processed. The following reagents will need to be purchased separately:

- [BL Buffer](#)

### **Materials and Reagents to be Supplied by User:**

- 15 mL Corex glass centrifuge tubes
- Centrifuge capable of 2,500g
- Vortexer
- 2.0 mL nuclease-free microcentrifuge tubes
- Nuclease-free pipette tips
- Water bath or heat block capable of 60°C and 70°C
- NaCl, EDTA, Tris-HCl, SDS,  $\beta$ -mercaptoethanol to prepare Buffer A and Buffer B (see table below in the Before Starting section)
- 100% ethanol
- 100% isopropanol
- Optional: 3M NaOH

### **Before Starting:**

- Heat the water bath or heat block to 60°C
- Heat Elution Buffer to 70°C
- Prepare the DNA Wash Buffer and HBC Buffer according to the instructions in the Preparing Reagents section on Page 6
- Prepare the following buffers before beginning:

Buffer A:	150 mM NaCl 10 mM EDTA, pH 8.0	Buffer B:	100 mM Tris-HCl, pH 8.0 10 mM EDTA 500 mM NaCl 1% SDS 2% $\beta$ -mercaptoethanol
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1. Add 50-250  $\mu\text{L}$  semen to 10 mL Buffer A in a 15 mL Corex centrifuge tube.  
**Note:** Using Corex tubes prevents attachment of the sperm cells to the tube walls.
2. Vortex for 10 seconds.
3. Centrifuge at 2,500g for 10 minutes.
4. Carefully aspirate the supernatant leaving  $\sim$ 1 mL pellet and Buffer A.
5. Vortex for 10 seconds.
6. Centrifuge at maximum speed to collect any sample adhering to the cap/walls of the tube.
7. Transfer sample to a new 2.0 mL microcentrifuge tube.
8. Add 500  $\mu\text{L}$  Buffer A to the Corex tube from **Step 4** to rinse the tube.
9. Vortex for 30 seconds.
10. Centrifuge at maximum speed to collect any sample adhering to the cap/walls of the tube.
11. Transfer sample from **Step 8** to the 2 mL nuclease-free microcentrifuge tube.
12. Centrifuge at maximum speed for 2 minutes.
13. Carefully remove the supernatant without disturbing the semen pellet.
14. Add 200  $\mu\text{L}$  Buffer B and resuspend pellet.

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15. Add 250  $\mu$ L BL Buffer.
  
16. Add 260  $\mu$ L 100% ethanol. Vortex to mix thoroughly.
  
17. Centrifuge at maximum speed to collect any sample adhering to the cap/walls of the tube.
  
18. Insert a HiBind<sup>®</sup> DNA Mini Column into a 2 mL Collection Tube.

## Optional Protocol for Column Equilibration:

1. Add 100  $\mu$ L 3M NaOH to the HiBind<sup>®</sup> DNA Mini Column.
  2. Let sit at room temperature 4 minutes.
  3. Centrifuge at maximum speed for 30 seconds.
  4. Discard the filtrate and reuse the collection tube.
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19. Transfer the entire sample from **Step 19** to the column, including any precipitate that may have formed.
  
  20. Centrifuge at maximum speed for 1 minute.
  
  21. Discard the filtrate and the collection tube.
  
  22. Transfer the HiBind<sup>®</sup> DNA Mini Column to a new 2 mL Collection Tube.
  
  23. Add 500  $\mu$ L HBC Buffer.  
  
**Note:** HBC Buffer must be diluted with 100% isopropanol before use. Please see Page 6 for instructions.
  
  24. Centrifuge at maximum speed for 1 minute.
  
  25. Discard the filtrate and the collection tube.

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26. Transfer the HiBind<sup>®</sup> DNA Mini Column to a new 2 mL Collection Tube.

27. Add 700  $\mu$ L DNA Wash Buffer.

**Note:** DNA Wash Buffer must be diluted with 100% ethanol before use. Please see Page 6 for instructions.

28. Centrifuge at maximum speed for 1 minute.

29. Discard the filtrate and reuse the collection tube.

30. Repeat Steps 27-29 for a second DNA Wash step.

31. Centrifuge at maximum speed for 2 minutes to dry the column.

**Note:** It is critical to remove all residual ethanol that might interfere with downstream applications.

32. Place the column into a 1.5 mL or 2.0 mL nuclease-free microcentrifuge tube.

33. Add 100  $\mu$ L Elution Buffer heated to 70°C.

34. Let sit at room temperature for 3 minutes.

35. Centrifuge at maximum speed for 1 minute.

36. Repeat Steps 35-37 for a second elution step.

**Note:** Incubation at 70°C will give a modest increase in DNA yield per elution. Using the first eluate for the second elution will increase DNA concentration.

37. Store DNA at -20°C.

## Troubleshooting Guide

Please use this guide to troubleshoot any problems that may arise. For further assistance, please contact the technical support staff, toll free, at **1-800-832-8896**.

Problem	Cause	Solution
Clogged Column	Incomplete Lysis	Extend incubation time with BL Buffer and Proteinase K Solution.
	Too much Sample	Divide sample into multiple tubes and adjust the volume to 250 $\mu$ L with BL Buffer.
	Sample is Viscous	Divide sample into multiple tubes and adjust the volume to 250 $\mu$ L with BL Buffer.
Problem	Cause	Solution
Low DNA Yield	Poor Elution	Repeat elution with increased elution volume. Incubate columns at 65°C for 5 minutes with Elution Buffer.
	Improper Washing	DNA Wash Buffer must be diluted with 100% ethanol before use. If refrigerated, DNA Wash Buffer must be brought to room temperature.
		HBC Buffer must be diluted with isopropanol before use. If refrigerated, HBC Buffer must be brought to room temperature.
	Sample has low DNA Content	Increase starting material and volume of all reagents (Proteinase K Solution, BL Buffer, ethanol) proportionally. Load aliquots of lysate through column successively.
Prime Columns	Add 100 $\mu$ L 3M NaOH to the column prior to loading the sample. Let sit for 4 minutes. Centrifuge at 10,000g for 60 seconds. Add 100 $\mu$ L water to the column and centrifuge at 10,000g for 60 seconds. Discard the filtrate.	



For more purification solutions, visit [www.omegabiotek.com](http://www.omegabiotek.com)

## AVAILABLE FORMATS



Spin Columns



96-Well Silica Plates



Mag Beads

## SAMPLE TYPES



Blood / Plasma



Plasmid



Cultured Cells



Plant & Soil



NGS Clean Up



Tissue




FFPE





Fecal Matter



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