E.Z.N.A.® Cycle Pure Kit

Revision No: 2.2

omega BIO-TEK

Quick Guide

Please visit www.omegabiotek.com for a downloadable user manual containing additional protocols, troubleshooting tips, and ordering information.

Product	D6492-00	D6492-01	D6492-02	D6492-03
Purifications	5	50	200	600
HiBind® DNA Mini Columns	5	50	200	600
2 mL Collection Tubes	5	50	200	600
CP Buffer	5 mL	40 mL	150 mL	3 x120 mL
DNA Wash Buffer	2.5 mL	25 mL	3 x 25 mL	200 mL
Elution Buffer	15 mL	30 mL	30 mL	35 mL

Supplied by user:

- Tabletop microcentrifuge capable of 13,000*g*
- Vortexer
- Nuclease-free 1.5 mL microcentrifuge tubes
- 100% ethanol
- For fragments <200 bp, 100% isopropanol
- Optional: sterile deionized water or TE Buffer
- Optional: Compatible vacuum manifold

Before starting:

- Prepare DNA Wash Buffer according to the directions on the bottle.

DNA Purification of PCR Products - Centrifugation Protocol

BIND

1. Perform agarose gel/ethidium bromide electrophoresis to analyze PCR product.
2. Determine the volume of your PCR reaction.
3. Transfer the sample into a clean 1.5 mL microcentrifuge tube (not provided).
4. Add 4-5 volumes CP Buffer. For PCR products smaller than 200 bp, add 5 volumes CP Buffer and 0.4 volumes 100% isopropanol.

   Note: Volume refers to the size of your PCR reaction. For example, if your PCR reaction is 100 µL and is smaller than 200 bp, you would use 500 µL CP Buffer and 40 µL isopropanol.
5. Vortex to mix thoroughly. Briefly centrifuge to collect any drops from the inside of the lid.
6. Insert a HiBind® DNA Mini Column into a 2 mL Collection Tube.
7. Add the sample from Step 5 to the HiBind® DNA Mini Column.

WASH

8. Centrifuge at maximum speed (≥13,000*g*) for 60 seconds at room temperature. Discard the filtrate and reuse the collection tube.
9. Add 700 µL DNA Wash Buffer diluted with 100% ethanol (see the bottle for instructions). Centrifuge at maximum speed for 60 seconds. Discard the filtrate and reuse the collection tube.
10. Repeat Step 9 for a second DNA Wash Buffer Step.

ELUTE

11. Centrifuge the empty HiBind® DNA Mini Column at maximum speed for 2 minutes to dry the column. This step is critical for removal of trace ethanol that may interfere with downstream applications.
12. Transfer the HiBind® DNA Mini Column into a clean 1.5 mL microcentrifuge tube.
13. Add 30-50 µL Elution Buffer, TE Buffer, or sterile deionized water directly to the center of column matrix. Let sit at room temperature for 2 minutes. Centrifuge at maximum speed for 60 seconds.
14. Store DNA at -20°C.

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Quick Guide

DNA Purification of PCR Products – Vacuum Protocol

BIND

1. Perform agarose gel/ethidium bromide electrophoresis to analyze PCR product.
2. Determine the volume of your PCR reaction.
3. Transfer the sample into a clean 1.5 mL microcentrifuge tube (not provided).
4. Add 4-5 volumes CP Buffer. For PCR products smaller than 200 bp, add 5 volumes CP Buffer and 0.4 volumes 100% isopropanol.

   Note: Volume refers to the size of your PCR reaction. For example, if your PCR reaction is 100 µL and is smaller than 200 bp, you would use 500 µL CP Buffer and 40 µL isopropanol.
5. Vortex to mix thoroughly. Briefly centrifuge to collect any drops from the inside of the lid.
6. Prepare the vacuum manifold according to manufacturer's instructions and connect the HiBind® DNA Mini Column to the manifold.

WASH

7. Transfer the entire sample to the HiBind® DNA Mini Column. Switch on the vacuum source to draw the sample through the column. Turn off the vacuum.
8. Add 700 µL DNA Wash Buffer diluted with 100% ethanol (see the bottle for instructions). Switch on the vacuum source to draw the DNA Wash Buffer through the column. Turn off the vacuum.
9. Repeat Step 8 for a second DNA Wash Buffer Step.
10. Transfer the HiBind® DNA Mini Column into a 2 mL Collection Tube.
11. Centrifuge the empty HiBind® DNA Mini Column at maximum speed (≥13,000*g*) for 2 minutes to dry the column. This step is critical for removal of trace ethanol that may interfere with downstream applications.

ELUTE

12. Transfer the HiBind® DNA Mini Column into a clean 1.5 mL microcentrifuge tube.
13. Add 30-50 µL Elution Buffer, TE Buffer, or sterile deionized water directly to the center of column matrix. Let sit at room temperature for 2 minutes. Centrifuge at maximum speed for 60 seconds.
14. Store DNA at -20°C.